

Introduction to Protein – DNA Interactions

Structure, Thermodynamics, and Bioinformatics

ALSO FROM COLD SPRING HARBOR LABORATORY PRESS

Other Titles of Interest

Bioinformatics: Sequence and Genome Analysis, Second Edition

Genes & Signals

A Genetic Switch, Third Edition: *Phage Lambda Revisited*

Molecular Cloning: A Laboratory Manual, Fourth Edition

Introduction to Protein–DNA Interactions

Structure, Thermodynamics, and Bioinformatics

GARY D. STORMO, PH.D.



COLD SPRING HARBOR LABORATORY PRESS
Cold Spring Harbor, New York • www.cshlpress.org

Introduction to Protein–DNA Interactions

Structure, Thermodynamics, and Bioinformatics

© 2013 by Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York

All rights reserved

Printed in the United States of America

<i>Publisher</i>	John Inglis
<i>Acquisition Editors</i>	Ann Boyle and Kaaren Janssen
<i>Director of Editorial Development</i>	Jan Argentine
<i>Developmental Editor</i>	Judy Cuddihy
<i>Project Manager</i>	Maryliz Dickerson
<i>Permissions Coordinator</i>	Carol Brown
<i>Production Manager</i>	Denise Weiss
<i>Production Editor</i>	Rena Steuer
<i>Compositor</i>	Techset Ltd.
<i>Cover Designer</i>	Ed Atkeson

Front cover: Computer-generated structural diagram showing the overall geometry of the Lac repressor protein binding to *lac* operator DNA (image generated using Pymol software from data in the Protein Data Bank database, entry 2KEI).

Library of Congress Cataloging-in-Publication Data

Stormo, Gary.

Introduction to protein-DNA interactions : structure, thermodynamics, and bioinformatics / Gary D. Stormo.

p. ; cm.

Includes bibliographical references and index.

ISBN 978-1-936113-49-1 (hard cover : alk. paper) – ISBN 978-1-936113-50-7 (pbk. : alk. paper)

I. Title.

[DNLM: 1. DNA-Binding Proteins–pharmacokinetics. 2. Binding Sites. 3. DNA–chemistry. 4. Protein Binding. 5. Transcription Factors. QU 58.5]

572.8'6459–dc23

2012035448

10 9 8 7 6 5 4 3 2 1

All World Wide Web addresses are accurate to the best of our knowledge at the time of printing.

Authorization to photocopy items for internal or personal use, or the internal or personal use of specific clients, is granted by Cold Spring Harbor Laboratory Press, provided that the appropriate fee is paid directly to the Copyright Clearance Center (CCC). Write or call CCC at 222 Rosewood Drive, Danvers, MA 01923 (508-750-8400) for information about fees and regulations. Prior to photocopying items for educational classroom use, contact CCC at the above address. Additional information on CCC can be obtained at CCC Online at <http://www.copyright.com>.

For a complete catalog of Cold Spring Harbor Laboratory Press publications, visit our website at www.cshlpress.org.

*To my parents, Milo and Claryce, who gave me the love
of learning and the encouragement to follow wherever that led.*

*To my wife, Susan Dutcher, and my children, Ben and Adrienne,
who have enriched my life immeasurably.*



Contents

Preface, ix

1 Importance of Protein–DNA Interactions, 1

STRUCTURE

2 The Structure of DNA, 13

3 Protein Structure and DNA Recognition, 27

4 Sequence-Specific Interactions in Protein–DNA Complexes, 49

THERMODYNAMICS

5 Binding Affinity, Cooperativity, and Specificity, 67

6 Energetics and Kinetics of Binding, 89

BIOINFORMATICS

7 Bioinformatics of DNA-Binding Sites, 109

8 Bioinformatics of Transcription Factors and Recognition Models, 131

9 Transcriptional Genomics, 153

Index, 193

Preface

THE BIOLOGICAL IMPORTANCE of PROTEIN–DNA INTERACTIONS has been recognized since the early 1960s, starting with the discovery by Jacob and Monod of the *lac* operon and its regulation in *Escherichia coli*. In the intervening 50 years, studies of protein–DNA interactions have made significant contributions to most areas of molecular, cellular, and developmental biology. A wide range of approaches has been applied in those studies, but they can be broadly classified into the three types that are the focus of this book: structural, thermodynamic, and bioinformatic. The earliest studies used biochemical and biophysical methods to analyze the thermodynamic and kinetic aspects of protein–DNA interactions. The first binding site sequences were determined in the early 1970s, which led to hypotheses about recognition mechanisms and the information required for regulatory systems to function. Technological advances in the late 1970s and the early 1980s, including the ability to sequence and synthesize DNA and to clone, express, and purify large quantities of proteins, facilitated many new types of studies. The earliest bioinformatics approaches were developed in the late 1970s, as soon as there were enough sequences for statistical analyses to be worthwhile. Shortly after that, as it became much easier to synthesize and purify sufficient quantities of specific proteins and DNA sequences of interest, structural studies rapidly increased. Further technological advances in the last two decades have continued to accelerate the pace of discovery. Most important have been further efficiencies in DNA sequencing that have resulted not only in whole-genome sequences for many species but also whole-genome and mRNA sequences from individuals as well as a variety of other sequence-based data sets. Our understanding of protein–DNA interactions and their roles in a wide range of biological processes has grown enormously, but there is still much we do not know and the field continues to be ripe for further discovery.

The primary goal of this book is to provide an introduction to protein–DNA interactions that bridges the three classes of approaches. Experts in any of the fields are not

likely to learn anything new within their field; in fact, they will undoubtedly find examples of details being glossed over in favor of a simplified presentation. But experts in one area tend to have more cursory knowledge of the other fields and thus may learn from other sections of the book. Those who are new to the study of protein–DNA interactions or those outside the field with a casual interest in the topic may gain new insights throughout the book. If so, the book has succeeded even beyond the fact that I learned something in the process of writing every chapter.

The regulation of gene expression has fascinated me since my graduate school days. I have ventured into other topics, mostly related to how computer programs can help to uncover biological knowledge, but the majority of my efforts have been focused on understanding how networks of transcription factors regulate gene expression and control cell fates and phenotypes. I have been extremely fortunate to have been associated throughout my career with teachers and students, colleagues and collaborators, and most of all friends who have taught and encouraged me and made my whole adventure enjoyable. The list of those who made significant contributions to my research, many of whom I have never met but have benefited from immensely through reading their papers, is too long to include in this preface. But a few have had such a large influence that I must thank them here. Larry Gold, my graduate and postdoc advisor, kept research always fun and gave me the freedom and encouragement to follow an unconventional path. Tom Schneider, a fellow student in Larry’s lab, and Andrej Ehrenfeucht, a mentor in all things computational, were there from the beginning and opened my eyes to new horizons that I would have missed without them. I have had many great collaborators over the years but special thanks go to John Heumann, Alan Lapedes, and Charles “Chip” Lawrence, each of whom has filled gaps in my knowledge and provided numerous insights into my own work that were initially invisible to me. I have also had many great students and postdocs who made progress possible and who taught me at least as much as I taught them.

This book would not have happened with the support and encouragement of the individuals at Cold Spring Harbor Laboratory Press, including Ann Boyle, Maryliz Dickerson, Kaaren Janssen, and Rena Steuer. Judy Cuddihy, in particular, made numerous improvements and helped at every step. I also thank those authors and publishers who allowed me to use their figures.

Index

A

- Adenine (A), 13
- A-DNA, 24–26
- Affinity, binding. See Binding affinity
- α helix secondary structure, 32–33
- AMBER, 101
- Amino acid properties
 - categories, 29, 30f
 - nonpolar hydrophobic amino acids, 29, 31
 - polar acidic residues, 32
 - polar basic residues, 31
 - polar uncharged residues, 31
 - special cases, 32
 - structural function, 29
- Artemisinic acid, 167
- Association constant (K_A), 69, 98. See also Binding affinity
- Avery, Oswald, 14

B

- B1H (bacterial one-hybrid) methods, 84
- Bacillus* genus, 163, 164
- Bacteriophage λ
 - choice between lysis and lysogeny, 5, 7–8
 - competition for binding between Cro and λ , 7
 - operator binding sites, 7
 - principles of sequence-specific TFs, 8
 - regulatory region elements, 5–6
- Basic-region leucine zipper (bZIP), 41, 95, 144–145
- B-DNA, 24–26
- Beadle, George, 18
- bHLH (basic-region helix-loop-helix), 41, 95, 144–145
- Binding affinity
 - assay methods, 76
 - binding probability equation, 69
 - determination methods, 79–80

- factors influencing the rate of complex formation, 68, 70f
- K_D determination methods, 70f
 - about, 69
 - EMSAs, 71–72
 - filter-binding assays, 71
 - fluorescence anisotropy, 73
 - SPR, 72–73
- measuring affinities of multiple sites simultaneously, 80–81
- nonspecific, 49
- Binding cooperativity
 - about, 73–74
 - affinity assay methods, 76
 - cooperativity constant, 74
 - nuclease protection, 76–78
 - physical basis of positive cooperativity, 75–76
 - probability-of-each-state calculations, 74–75
- Binding location analyses, 160–161
- Binding-site motifs discovery
 - expectation maximization, 126–127
 - Gibbs sampling, 127–128
 - greedy alignments, 125–126
 - “motif discovery” problem, 123, 124f
 - pattern searches, 123–125
 - pros and cons of methods, 123
- Binding specificity
 - about, 78
 - bioinformatics of DNA-binding sites and, 109–110
 - estimating specificity needed for a regulatory system, 78–79
 - limits to specificity determination, 149
 - methods for determining
 - bacterial one-hybrid, 84
 - basis of, 80
 - CSI, 83
 - determining affinity and, 79–80

Binding specificity (*Continued*)
 measuring affinities of multiple sites
 simultaneously, 80–81
 MITOMI, 81–83
 PBMs, 83, 148
 SELEX, 83–84, 148
 quantitative definition of specificity, 84–86
 recognition model used to determine, 147–148
 sequence-specific interactions (See
 Sequence-specific interactions)
 specificity modeling by PWM
 discriminatory models, 119
 higher-order models, 121–122
 probabilistic models, 113–119
 regression models, 119–121
 Bioinformatics of DNA-binding sites
 position weight matrix (See Position weight
 matrix)
 representation of the specificity of TFs, 109–110
 Bioinformatics of TFs and recognition models
 hidden Markov model
 examples of TF profile HMMs, 143–146
 probability of generating a particular
 sequence, 142–143
 protein sequences alignment example,
 138–140
 pseudocounts additions, 141
 sequence logos, 141–142
 types of states, 140–141
 identifying homologous TFs
 assessing if two proteins are homologous,
 134
 BLAST database search method, 134, 138
 BLOSUM62 substitution matrix, 132–134
 methods used to predict the function of a
 protein, 131
 mutations and, 132
 optimal alignments with dynamic program-
 ming, 135–137
 orthologs and paralogs, 132
 recognition models
 binding specificity determination method,
 147–148
 focus on developing a predictive model,
 146–147
 lack of a recognition code and, 146
 limitations of the recognition code, 149–150
 limits to specificity determination, 149
 method to determine binding specificity,
 147–148
 phage-display method, 148–149
 quantitative models, 150
 BLAST database search method, 136, 138
 BLOSUM62 substitution matrix, 132–134
 Britten, Roy, 15
 β strands and β sheets secondary structure, 33–34
 bZIP (basic-region leucine zipper), 41, 95, 144–145

C

C2H2 zinc finger family, 138, 140–143, 144
Caenorhabditis elegans, 159
 “Calling cards,” 160–161
 cAMP (cyclic AMP), 4, 5
 cAMP receptor protein (CRP), 4, 94, 123, 156
 Cancer Genome Anatomy Project (CGAP), 174
 CAP (catabolite activator protein), 4
 Carroll, Sean, 173
 CGAP (Cancer Genome Anatomy Project), 174
 CHARMM, 101
 ChIP (chromatin immunoprecipitation), 123, 160–161
 ChIP-seq experiments, 160, 162, 168–170
 chip-seq experiments, 175
 Cognate site identifier (CSI), 83
 Coiled-coil helix dimers, 41, 42f
 Cooperativity. See Binding cooperativity
 COSY (correlation spectroscopy), 38
 Crick, Francis H.C., 13
 Cro protein, 5–8, 35–36
 CRP (cAMP receptor protein), 4, 94, 123, 156
 Cyclic AMP (cAMP), 4, 5
 Cytosine (C), 13

D

DamID, 160–161
 Delete states, 140–141
 DHS (DNase I hypersensitive sites), 162, 170–172
 Discriminatory models for specificity, 119
 Dissociation constant (K_D), 70f. See also Binding affinity
 about, 69
 EMSAs, 71–72
 filter-binding assays, 71
 fluorescence anisotropy, 73
 SPR, 72–73
 DNA accessibility analyses, 162
 DNase I, 162
 DNase I hypersensitive sites (DHS), 162, 170–172
 Double helix. See Structure of DNA
Drosophila
 conservation of enhancers function, 181–182,
 183f
 embryonic development steps, 179–180
 history as a model organism, 177
 research advances, 180
 dsDNA (double-stranded DNA), 15–17
 Dynamic programming, 135–137

E

EcoCyc, 163
 EcoRI, 53–54
 Eggert, M, 125
 EM (expectation maximization), 126–127
 EMSAs (electrophoretic mobility-shift assays), 71–72
 ENCODE project
 about, 167–168
 challenges in studying multicellular eukaryotes,
 167

ChIP-seq experiments, 168–170
 DNase I hypersensitive experiments,
 170–172
 project expansion, 168, 169f
endo16 gene, 177, 179f
 Energetics and kinetics of binding. See Thermody-
 namics of TF binding
 Enhancers, 9
 Enthalpy (H), 90, 92, 94–95
 Entropy (S), 90, 92–95, 117, 119
Escherichia coli
 gene expression regulation and the *lac* operon,
 3–5
 gene regulatory networks study, 163–165
 scaling up to human dimensions example,
 17–18
even-skipped (*eve*), 180, 181, 182f, 183f
 Expectation maximization (EM), 126–127
 Expression analyses, 157–160

F

FAIRE (formaldehyde-assisted isolation of regulatory
 elements), 162
 FFL (feed-forward loop), 156–157
 Filter-binding assays, 71
 Fluorescence anisotropy, 73
 Fly-Ex, 180
 FlyNet, 180
 “Fly Room” laboratory, 177
 Formaldehyde-assisted isolation of regulatory elements
 (FAIRE), 162
fushi tarazu (*ftz*), 180

G

Galas, DJ, 125
 GATA family, 43–44, 143–144
 Gel- or band-shift assays, 71–72
 Gene expression regulation. See also Gene regulatory
 networks
 bacteriophage λ
 choice between lysis and lysogeny, 7–8
 competition for binding between Cro
 and λ , 7
 operator binding sites, 7
 principles of sequence-specific TFs, 8
 regulatory region elements, 5–6
lac operon of *E. coli* and, 3–5
 mystery of, 2–3
 principles of protein–DNA interactions and,
 18–19
 specificity of TFs and, 78
 Generative probabilistic models, 115, 117
 Gene regulatory networks (GRNs)
 Binding-site information and, 157
 characteristics of biological networks, 156
Drosophila embryonic patterning, 177,
 179–182, 183f

feed-forward loop network motif, 156–157
 genetic variation and, 172–175
 modeling conventions, 154–155
 model systems’ characteristics, 175–176
 sea urchin studies, 176–177, 178f, 179f
 study of
 bacteria based, 163–165
 ENCODE project, 167–172
 genetic variation, 172–175
 limitations from studying only TFs and their
 targets, 162–163
 synthetic biology, 165–166
 yeast, 166–167
 “wiring diagram” uses, 155–156
 Genetic variation and GRNs
 concept of the “human genome,” 173–174
 genome-wide association studies, 173–174
 levels of DNA variation, 172–173
 regulation differences focus, 173
 sequence differences mechanisms focus, 173
 Genome-wide association studies (GWAS),
 174–175
 Gibbs sampling, 127–128
 Gibbs standard free energy of binding, 69, 89–90
 Greedy alignments, 125–126
 GRNs. See Gene regulatory networks
 Guanine (G), 13
 GWAS (Genome-wide association studies), 174–175

H

H (enthalpy), 90, 92, 94–95
 HapMap project, 174
 Helix-turn-helix protein family, 5, 35, 39–41,
 145–146
 Helix-turn-helix proteins, 51
 Hidden Markov model (HMM)
 examples of TF profile HMMs, 143–146
 probability of generating a particular sequence,
 142–143
 protein sequences alignment example,
 138–140
 pseudocounts additions, 141
 sequence logos, 141–142
 types of states, 140–141
 Higher-order models for specificity, 121–122
 Homeodomain proteins, 41
 Homodimers, 22
 Homologous TFs
 assessing if two proteins are homologous, 134
 BLAST database search method, 134, 138
 BLOSUM62 substitution matrix, 132–134
 methods used to predict the function of a
 protein, 131
 mutations and, 132
 optimal alignments with dynamic program-
 ming, 135–137
 orthologs and paralogs, 132

Human Microbiome Project, 164
Hydrophobic effect, 95

I

IC (information content) measurement, 118–119
Insert states, 140–141
International Genetically Engineered Machine (iGEM) Foundation, 165
Int protein, 7
Introns, 9
ITC (isothermal titration calorimetry), 92, 93

J

Jacob, François, 3

K

K_A (association constant), 69, 98. *See also* Binding affinity
 K_D . *See* Dissociation constant (K_D)
Kendrew, John, 36
King, Mary-Claire, 173
Kullback-Leibler distance, 119

L

lac operon of *E. coli*
 compared to the λ repressor, 8, 9
 gene expression regulation and, 3–5, 61
Lac repressor
 binding specificity of, 99, 103, 156
 helix-turn-helix protein family, 39–40
 lactose regulatory system and, 3–5, 8, 47
 sequence-specific interactions, 61–63
Lactose, 3–5, 47
Lewis, Edward, 177
Likelihood ratios, 116–117
Log-odds PWM, 117
 λ repressor protein, 5–8
Lysis/lysogeny decision of phage DNA, 5, 7–8

M

Major groove, 19–20, 51
Markov chain Monte Carlo (MCMC), 101
Match states, 140–141
MC (Monte Carlo) methods, 101
MD (molecular dynamics) simulations, 101
Melting DNA, 15–17
MicrobesOnline, 164
Minor groove, 20–21
MITOMI (mechanically induced trapping of molecular interactions), 81–83
Molecular dynamics (MD) simulations, 101
Monod, Jacques, 3
Monte Carlo (MC) methods, 101
Morgan, T.H., 177
Motif discovery problem, 123, 124f
mRNA (messenger RNA)

 measuring using microarrays, 157–158
 protein–DNA interactions and, 2, 3–5, 8–9
 role within a cell, 17
 sequencing, 159
Mullis, Dary, 16
Mutations and homologous TFs, 132

N

Ndt80, 59–60
NFAT (nuclear factor of activated T cells), 45
NF- κ B, 45
NMR (nuclear magnetic resonance), 37–39
NOESY (nuclear Overhauser effect spectroscopy), 38
Noncoding DNA, 9
Nonpolar hydrophobic amino acids, 29, 31
Nonspecific binding affinity, 49
Nuclear factor of activated T cells (NFAT), 45
Nuclear magnetic resonance (NMR), 37–39
Nuclear Overhauser effect spectroscopy (NOESY), 38
Nuclease protection, 76–78
Nucleosomes, 9
Nüsslein-Volhard, Christiane, 177

O

1D (one-dimensional) diffusion, 103
1000 Genomes Project, 174
One-dimensional (1D) diffusion, 103
Orthologs, 132

P

p53, 45
Paralogs, 132
Pauling, L., 172
PBMs (protein-binding microarrays), 83, 148
PCR (polymerase chain reaction), 16–17, 36
Perutz, Max, 36
PFM (position frequency matrix), 142
Phage display, 148–149
Phosphorylation, 46
Phylogenetic footprinting, 128, 129f
Polar acidic residues, 32
Polar basic residues, 31
Polar uncharged residues, 31
Polymerase chain reaction (PCR), 16–17, 36
Position frequency matrix (PFM), 142
Position weight matrix (PWM)
 advantages of, 111–112
 discovery of binding-site motifs
 expectation maximization, 126–127
 Gibbs sampling, 127–128
 greedy alignments, 125–126
 “motif discovery” problem, 123, 124f
 pattern searches, 123–125
 pros and cons of methods, 123
 phylogenetic footprinting, 128, 129f
 sequence and functional modeling using, 112–113

- specificity modeling
 discriminatory models, 119
 higher-order models, 121–122
 probabilistic models, 113–119
 regression models, 119–121
 uses, 110–111
- Probabilistic models for specificity
 generative model, 115, 117
 information content measurement, 118–119
 known binding sites basis, 113–115
 likelihood ratios and information content,
 116–117
- Profile HMM. See Hidden Markov model
- Promoters, 4
- Protein-binding microarrays (PBMs), 83, 148
- Protein cleavage, 46
- Protein–DNA complexes. See Protein structure;
 Sequence-specific interactions
- Protein–DNA interactions
 accessibility of genomic DNA, 9
 action-at-a-distance rule for eukaryotes, 9
 approaches to the study of, 10–11
 division of labor between proteins and DNA,
 1, 2
 functions performed by proteins on DNA, 1–2
 messenger RNA and, 2
 regulation of gene expression
 bacteriophage λ , 5–8
lac operon of *E. coli* and, 3–5
 mystery of, 2–3
 TFs and eukaryotic gene regulation, 9–10
 TFs in prokaryotes versus eukaryotes, 8–9
 transcription factors and, 2
- Protein structure
 allosteric effectors, 47
 amino acid properties
 nonpolar hydrophobic amino acids, 29,
 30f, 31
 polar acidic residues, 32
 polar basic residues, 31
 polar uncharged residues, 30f, 31
 side-chain categories, 29, 30f
 special cases, 32
 structural function, 29, 30f
- β strands and β sheets secondary structure,
 33–34
- determination methods, 36–39
- families
 classifications, 35
 coiled-coil helix dimers, 41, 42f
 helix-turn-helix proteins, 35, 39–41
 recognition with β strands, 44–45
 recognition with loops, 45
 zinc-coordinating proteins, 41–44
- functional domains, 34–35
- α helix secondary structure, 32–33
- levels, 27, 28f
- modifications, 46–47
- multi-protein complexes, 46
- protein–DNA complexes, 39–41
- protein sequence determination, 27–28
- PWM. See Position weight matrix
- ## R
- RAR (retinoic acid receptor), 42
- Recognition helix, 35, 41
- Recognition models
 binding specificity determination method,
 147–148
 with β strands, 44–45
 focus on developing a predictive model,
 146–147
 lack of a recognition code and, 146
 limitations of the recognition code, 149–150
 limits to specificity determination, 149
 with loops, 45
 phage display method, 148–149
 quantitative models, 150
 recognition code for zinc finger proteins,
 57–58
- Registry of standard biological parts, 165
- Regression models for specificity, 119–121
- Regtransbase, 164
- RegulonDB, 163
- Relative entropy, 117, 119
- Rel-homology domain, 45
- Retinoic acid receptor (RAR), 42
- Retinoid X receptor (RXR), 42
- Ribosomes, 2
- Romanuka, J., 61
- RTIDE, 125
- runt, 45
- RXR (retinoid X receptor), 42
- ## S
- S (entropy), 90, 92–95, 117, 119
- Saccharomyces cerevisiae*, 9, 166–167
- Saccharomyces* Genome database (SGD), 166
- Sarai, A, 80
- SBML (Systems Biology Markup Language), 156
- Sea urchin studies, 176–177, 178f, 179f
- Seeman, NC, 146
- SELEX (systematic evolution of ligands by exponential
 enrichment), 57, 83–84, 147, 148
- SELEX-seq, 148
- Sequence-specific interactions
 lessons on specificity of TFs, 64
 profiles of specificity
 EcoRI, 53–54
 Lac repressor, 61–63
 Ndt80, 59–60
 zinc finger proteins, 54f, 55–59
 specificity of protein–DNA interfaces, 50–52
 specificity's meanings, 49–50

- Sequence-specific interactions (*Continued*)
 structures of nonspecific binding, 63–64
 σ factors (sequence-specific binding proteins), 164
 SGD (*Saccharomyces* Genome database), 166
 Simple consensus sequence, 109–110
 Single-stranded DNA (ssDNA), 15–17
 Smith, Michael, 17
 Smith–Waterman algorithm, 134, 137
 SNP variants, 175
 spbase database, 176
 Specificity, binding. *See* Binding specificity
 SPR (surface plasmon resonance), 72–73
 ssDNA (single-stranded DNA), 15–17
 STAT factors, 45
Strongylocentrotus purpuratus (sea urchin),
 176–177, 178f, 179f
 Structure of DNA
 accessible surfaces of base pairs, 19–21
 alternative structures, 24–26
 base pairs, 13–14
 DNA melting, 15–17
 implications of, 18
 major groove, 19–20
 minor groove, 20–21
 modified bases, 22
 potential symmetry of DNA sequences, 21–22
 principles of protein–DNA interactions and
 gene regulation, 18–19
 scaling *E. coli* up to human dimensions, 17–18
 sequence-dependent variation, 22–23
 Surface plasmon resonance (SPR), 72–73
 Synthetic biology, 165–166
 Systematic evolution of ligands by exponential enrichment (SELEX), 57, 83–84, 147, 148
 Systems Biology Markup Language (SBML), 156
- T**
- Takeda, Y, 80
 TAL (transcription activator-like), 45
 TATA-binding protein (TBP), 44, 95
 Tatum, Edward, 18
 TFs. *See* Transcription factors
 Thermodynamics of TF binding
 computational modeling, 100–102
 contributions of entropy and enthalpy, 94–95
 enthalpy of an interaction, 92
 entropy change in an interaction, 92–94
 free energy equation, 89–92
 heat capacity changes, 95–96, 97f
 kinetics of binding-site location, 102–105
 molecular contributions to complex formation
 direct and indirect readout, 100
 electrostatic and nonelectrostatic contributions, 98–99
 nature of interactions, 96, 98
 specific and nonspecific contributions,
 99–100
 Thymine (T), 13
 Transcription activator-like (TAL), 45
 Transcriptional genomics
 binding location analyses, 160–161
 conclusions, 183–184
 developments in DNA studies, 153–154
 DNA accessibility analyses, 162
 expression analyses, 157–160
 gene regulatory networks (*See* Gene regulatory
 networks)
 Transcription factors (TFs)
 allosteric effectors, 47
 families
 classifications, 35
 coiled-coil helix dimers, 41, 42f
 helix-turn-helix proteins, 5, 35, 39–41
 recognition with β strands, 44–45
 recognition with loops, 45
 function, 2
 functional domains, 34–35
 modifications, 46–47
 multiprotein complexes, 46
 Tryptophan, 47
- V**
- Van der Waals contacts, 19, 20
- W**
- Waterman, MS, 125
 Watson, James D., 13
 WEEDER, 125
 Wieschaus, Eric, 177
 Wilson, Allan, 173
 Winged HTH subfamily, 40–41
 Wolfe, S.A., 57
 Wüthrich, Kurt, 37
- X**
- X-ray crystallography, 36–37
- Z**
- Z-DNA, 25
 Zif268, 55–56, 58–59
 Zinc cluster, 42–43
 Zinc finger domain, 34, 41–42
 Zinc finger proteins, 54f, 55–59, 146–147
 Zuckerkandl, E, 172